



Quick-DNA/RNA[™] Viral Kit

Viral DNA & RNA from any biological sample

Highlights

- Quick, spin-column purification of viral DNA and RNA from plasma, ٠ serum, urine, cell culture media, blood, saliva, cellular suspensions, swab, fecal and biopsy samples
- High-guality DNA/RNA is ready for Next-Gen sequencing, RT/gPCR, hybridization, etc.
- DNA/RNA Shield is included for sample collection, inactivation, • storage and preservation.

Catalog Numbers: D7020, D7021



Scan with your smart-phone camera to view the online protocol/video.







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Product Contents

<i>Quick</i> -DNA/RNA [™] Viral Kit	D7020 (50 prep)	D7021 (200 prep)
DNA/RNA Shield [™] (2X concentrate)	25 ml	125 ml
Viral DNA/RNA Buffer ¹	25 ml (x2)	100 ml (x2)
Viral Wash Buffer ² (concentrate)	6 ml (x2)	24 ml (x2)
DNase/RNase-Free Water	6 ml	6 ml (x2)
Zymo-Spin [™] IIC-XLR Columns	50	200
Collection Tubes	100	400
Instruction Manual	1 pc	1 pc

Storage Temperature - Store all kit components (i.e., buffers, columns) at room temperature (15-30°C). Before use:

1 Add beta-mercaptoethanol (β -Me; user provided) to 0.5% (v/v) *i.e.*, add 125 µl or 500 µl β -Me per 25 ml or 100 ml Viral DNA/RNA Buffer.

2 Add 24 ml of 100% ethanol (26 ml of 95% ethanol) to the 6 ml Viral Wash Buffer concentrate (D7020) or 192 ml of 100% ethanol (204 ml of 95% ethanol) to the 48 ml Viral Wash Buffer concentrate (D7021).

Specifications

 Sample Sources – ≤ 400 µl plasma, serum, saliva, swab, urine, cell culture media, blood, cellular suspension, fecal sample or ≤ 25 mg biopsy sample.

For samples in UTM[®]/VTM[®], PBS or saline, see Sample Preparation, page 5.

- Purity DNA/RNA is ready for Next-Gen Sequencing, RT/qPCR, etc.
- Binding Capacity 50 µg DNA/RNA (Zymo-Spin[™] IIC-XLR Columns).
- Elution Volume $\geq 50 \ \mu l$ DNase/RNase-Free Water.
- Equipment Needed (user provided) Beta-mercaptoethanol (b-Me), Ethanol (95-100%), Microcentrifuge.
- Materials (available separately) –

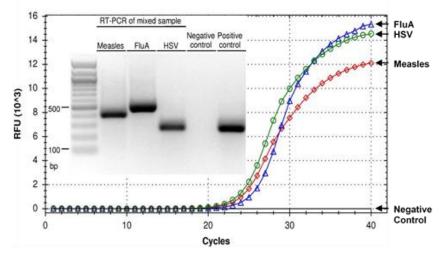
DNase I Set (E1010; 50 rxns.; 250 U DNase I (lyophilized) supplied w/ DNA Digestion Buffer, 4 ml) DNA/RNA Prep Buffer (D7010-2-50; 50 ml) DNA/RNA Wash Buffer (concentrate) (D7010-3-6, 6 ml) Proteinase K Set (D3001-2-20; 20 mg Proteinase K (lyophilized) supplied w/ Storage Buffer).

Product Description

The **Quick-DNA/RNA[™] Viral Kit** is a quick, purification of viral DNA and/or RNA from plasma, serum, urine, cell culture media, blood, saliva, cellular suspensions, biopsies, swab and fecal samples stored in **DNA/RNA Shield[™]** (for sample collection, nucleic acid preservation and inactivation of pathogens).

The kit also features a buffer system that facilitates complete viral particle lysis for efficient nucleic acid isolation. Small (> 50 nt) and large (> 200 kb) DNA and RNA are bound to the column, washed and eluted.

The isolated high-quality nucleic acids are ready for all downstream applications such as Next-Gen sequencing, hybridization-based and RT/qPCR detection.



Detection of DNA & RNA Viruses from a Mixed Population

Viral nucleic acids were isolated from liquid samples using the **Quick-DNA/RNA[™] Viral Kit**. Data shows RT-qPCR Ct values for measles, influenza type A (FluA), and herpes-simplex (HSV) viruses, 23.05 (diamonds), 24.56 (triangles), 22.92 (circles), respectively. Negative control – RT-PCR (no template w/ HSV specific primers). Positive control – PCR (HSV template w/ HSV primers).

Protocol

The protocol consists of: (I) Buffer Preparation, (II) Sample Preparation and (III) DNA/RNA Purification.

(I) Buffer Preparation

- Add beta-mercaptoethanol (user provided) to 0.5% (v/v) i.e., add 125 µl or 500 µl β-Me per 25 ml or 100 ml Viral DNA/RNA Buffer.
- ✓ Add 24 ml of 100% ethanol (26 ml of 95% ethanol) to the 6 ml Viral Wash Buffer concentrate (D7020) or 96 ml of 100% ethanol (104 ml of 95% ethanol) to the 24 ml Viral Wash Buffer concentrate (D7021).

(II) Sample Preparation

- ✓ Perform all steps at room temperature (15-30°C).
- ✓ Depending on sample type, up to 400 µl can be processed per prep (see below).

<u>Samples in DNA/RNA Shield^{™1,2} collection devices</u> (swabs, saliva, etc.) Transfer up to 400 µl and proceed directly with purification, page 6.

Swabs (UTM[®]/VTM[®], PBS, saline, etc.)

Transfer up to 400 µl and proceed directly with purification, page 6. Optional - To inactivate, store and preserve samples at room temperature prior to further processing, add **DNA/RNA Shield**[™]. See Liquids, below.

<u>Liquids</u> (plasma², serum², CSF, blood, saliva, urine, cell suspension, cell culture media) Add 200 µl of DNA/RNA Shield[™] (2X concentrate) to 200 µl liquid sample (1:1) and mix well. Transfer up to 400 µl of the mixture and proceed with purification, page 6.

<u>Tissue</u>² (LCM, needle biopsy)

Add 400 µl **DNA/RNA Shield**[™] (1X) to a tissue sample (up to 25 mg) and mix well. Proceed with purification, page 6.

Optional - **Proteinase K treatment**³ (protein-rich samples e.g., plasma, serum, saliva, sputum, tissue, can be treated). Materials sold separately.

Add 1% **Proteinase K** (v/v) at 20 mg/ml directly to a liquid sample. Mix well and incubate at room temperature for 15 minutes. Note: Up to 5% Proteinase K can be added (e.g., tissue). For example: Add 4-20 μ l Proteinase K to each 400 μ l sample.

¹ At this point, samples in DNA/RNA Shield[™] can be stored at ambient temperature (4-30°C) for a month, 7 days at 37°C, or long-term (> 1 year) -20°C or below.

² To remove particulate debris or cryoprecipitates (if any), centrifuge and transfer up to 400 µl of the cleared supernatant into a nuclease-free plate/tube (not provided).

³ Prior to use, reconstitute the lyophilized Proteinase K (D3001-2-20) and add 1,040 µl Storage Buffer. Mix well and store frozen aliquots.

(III) DNA/RNA Purification

- ✓ Perform all steps at room temperature and centrifugation at 10,000-16,000 x g.
- \checkmark The sample input can be scaled up or down, proportionally.
- 1. Add 800 μl **Viral DNA/RNA Buffer** to each 400 μl sample¹ (2:1) and mix well.
- Transfer the mixture into a Zymo-Spin[™] IIC-XLR Column² in a Collection Tube and centrifuge for 2 minutes. Transfer the column into a new collection tube.

Optional: At this point, DNase I treatment can be performed (see Appendices, page 7).

- 3. Add 500 µl **Viral Wash Buffer** to the column, centrifuge for 30 seconds and discard the flow-through. <u>Repeat this step</u>.
- Add 500 µl ethanol (95-100%) to the column and centrifuge for 1 minute to ensure complete removal of the wash buffer. Carefully, transfer the column into a nuclease-free tube (not provided).
- 5. Add 50 µl **DNase/RNase-Free Water** directly to the column matrix and centrifuge for 30 seconds.

Alternatively, for highly concentrated DNA/RNA use \geq 35 µl elution.

The eluted DNA/RNA³ can be used immediately or stored frozen.

¹ Up to 400 µl sample (including the volume of DNA/RNA Shield, if added) can be processed per prep.

² To process > 700 µl, the column can be reloaded.

³ It is recommended to titrate the DNA/RNA eluate for downstream applications (i.e., RT/qPCR, etc.).

Appendices

DNase I Treatment

✓ For DNA-free RNA, DNase I treatment can be performed using DNase I Set (E1010; 50 reactions), DNA/RNA Prep Buffer (D7010-2-50) and DNA/RNA Wash Buffer (concentrate) (D7010-3-6); materials sold separately.

For each sample to be treated, prepare **DNase I Reaction Mix** in an RNase-free tube (not provided) and mix by gentle inversion:

DNase I Reaction Mix	
DNA Digestion Buffer	75 µl
DNase I (reconstituted; 1 U/ul) ^{1,2}	5 µl

- Following DNA/RNA binding (page 6, step 2), add 400 μl DNA/RNA Wash Buffer³ to the column, centrifuge and discard the flow-through.
- 2. Add 80 µl DNase I Reaction Mix directly to the matrix of the column.
- 3. Incubate at room temperature for (15-30°C) for 15 minutes.
- 4. Add 500 µl **DNA/RNA Prep Buffer** to the column, centrifuge and discard the flow-through.
- 5. Proceed with DNA/RNA Purification (page 6, step 3).

¹ Prior to use, reconstitute lyophilized 250 U **DNase I** (E1009-A) to 1U/µl (final concentration) with 275 µl nuclease-free water (not provided), mix by gentle inversion and store frozen aliquots.

² Unit definition – one unit increases the absorbance of a high molecular weight DNA solution at a rate of 0.001 A260 units/ml of reaction mixture at 25°C.

³ Before use, add 24 ml of 100% ethanol (26 ml of 95% ethanol) to the 6 ml DNA/RNA Wash Buffer concentrate.

Ordering Information

Product Description	Catalog No.	Size
<i>Quick</i> -DNA/RNA [™] Viral Kit	D7020 D7021	50 preps. 200 preps.

Individual Kit Components	Catalog No.	Amount
DNA/RNA Shield [™] (2X concentrate)	R1200-25 R1200-125	25 ml 125 ml
Viral DNA/RNA Buffer	D7020-1-25 D7020-1-100	25 ml 100 ml
Viral Wash Buffer (concentrate)	R1034-2-24 R1034-2-48	24 ml 48 ml
Zymo-Spin™ IIC-XLR	C1104-25 C1104-50	25 50
Collection Tubes	C1001-50 C1001-500	50 500
DNase/RNase-Free Water	W1001-30 W1001-100	30 ml 100 ml
DNA/RNA Shield [™] Fecal Collection Tube	R1101	10
DNA/RNA Shield [™] Collection Tube DNA/RNA Shield [™] Lysis Tube (microbe) DNA/RNA Shield [™] Lysis Tube (microbe) w/ swab DNA/RNA Shield [™] Lysis Tube (tissue)	R1102 R1103 R1104 R1105	50 50 50 50
DNA/RNA Shield [™] Collection Tube w/ Swab (1 ml fill)	R1106 R1107	10 50
DNA/RNA Shield [™] Collection Tube w/ Swab (2 ml fill)	R1108 R1109	10 50
DNA/RNA Shield [™] Saliva Collection Kit (2 ml fill)	R1210	1
DNase I Set (250 U DNase I (lyophilized) supplied with DNA Digestion Buffer, 4 ml)	E1010	1
DNA/RNA Prep Buffer	D7010-2-50 D7010-2-200	50 ml 200 ml
DNA/RNA Wash Buffer	D7010-3-6 D7010-3-24	6 ml 24 ml
Proteinase K Set supplied w/ Storage Buffer	D3001-2-5 D3001-2-20	5 mg 20 mg

Complete Your Workflow

 ✓ For sample collection, inactivation of pathogens, storage and preservation of nucleic acids, use DNA/RNA Shield[™] collection devices:

DNA/RNA Shield [™] Collection Devices	
DNA/RNA Shield [™] Collection Tube w/ Swab (1 ml fill or 2 ml fill) #R1107, R1109	For swab samples of nasal, throat, etc.
DNA/RNA Shield [™] Saliva Collection Kit (2 ml fill) #R1210	For saliva, sputum, etc.
DNA/RNA Shield [™] Collection Tube DNA/RNA Shield [™] Lysis Tube (microbe) DNA/RNA Shield [™] Lysis Tube (microbe) w/ swab DNA/RNA Shield [™] Lysis Tube (tissue) #R1102-R1105	For microbes, tissue, etc. (2 ml lysis tubes used for bead beating homogenization)

✓ For RNA clean-up (purification) from the aqueous phase (e.g., TRIzol, TRI Reagent or similar) or from any enzymatic reaction (e.g., DNase I treated RNA):

RNA Clean & Concentrator	
Microprep #R1013, R1015	DNase I Set included (#R1013)
MagBeads #R1081, R1082	(#R1082)

Troubleshooting Guide

Problem	Possible Causes and Suggested Solutions
RNA degradation	To prevent RNA degradation: Immediately collect and lyse fresh sample into a stabilization reagent (i.e., DNA/RNA Shield [™]) to ensure nucleic acid stability. Homogenized samples in DNA/RNA Shield [™] can be stored frozen for later processing.
Low nucleic acid content and/or low sensitivity in downstream application	 Incomplete deproteinization due to high-protein content in the sample (blood, plasma/serum, tissue etc.): Increase the volume of DNA/RNA Shield[™] to the sample. Perform Proteinase K treatment (see Sample Preparation, page 4). Increase eluate input: Titrate the DNA/RNA eluate for downstream applications (i.e., RT/qPCR).
DNA contamination	To remove DNA: - Perform DNase I treatment during the purification (page 6) or perform DNase I treatment post-purification (#R1017), then clean-up the treated sample.

For technical assistance, please contact 1-888-882-9682 or email tech@zymoresearch.com

Notes

Notes



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